



Molecular Dynamics Simulations of Lysozyme in Sugar Aqueous Solutions

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ANR program « Physique-Chimie du Vivant »

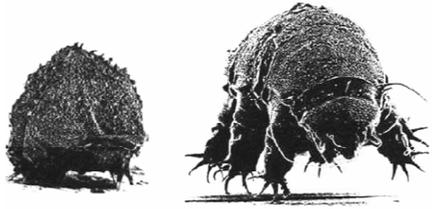
2007-2011 BIOSTAB project



Biopreservation

Anhydrobiosis

Some organisms can withstand extreme conditions of drought and/or temperature

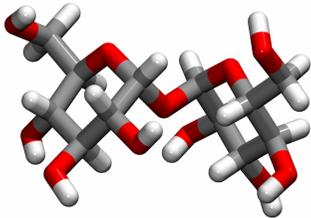


tardigrade

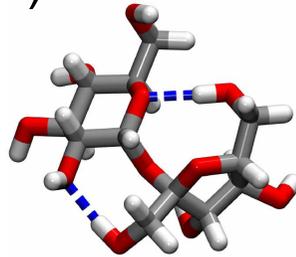


resurrection plant

They naturally synthesize sugars (trehalose, sucrose, etc.)



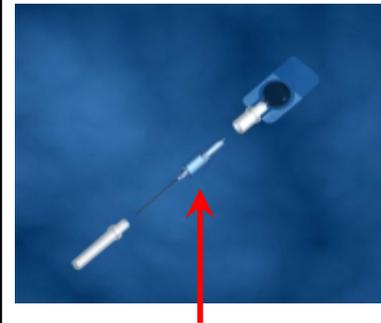
Trehalose



Sucrose

Lyophilisation (freeze-drying)

Long-term conservation of high-value products (therapeutic proteins, blood, organs, etc.)



Dried vaccine

potentially destructive process (ice formation, dehydration, pH and c° changes)



- ▶ Molecular mechanisms ?
- ▶ Why is trehalose more efficient ?

Hypotheses

- Glass formation
- T_{2H_2O}/T_{α} interconversion
- Water replacement
- Destructuring effect
- Water entrapment

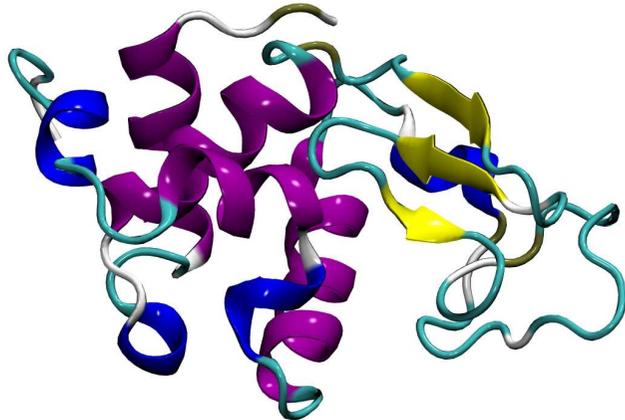


None fully satisfactory

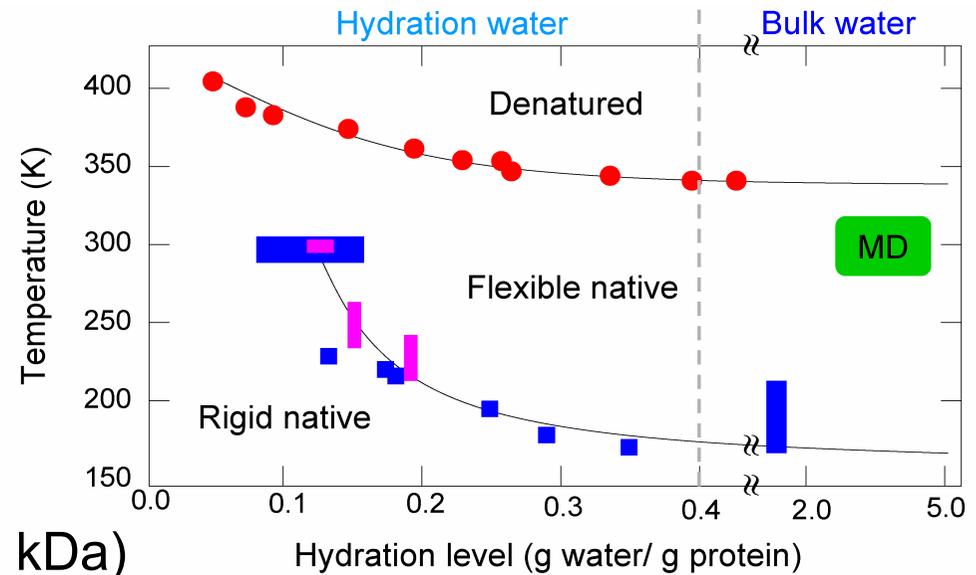
Simulation details

Model protein

Lysozyme (PDB : 193L)

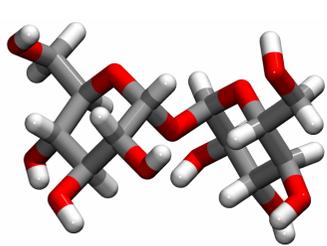


Small globular protein (129 residues, 14.3 kDa)

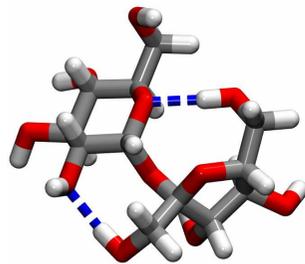


Gregory, in « The Properties of Water in Foods », 1997

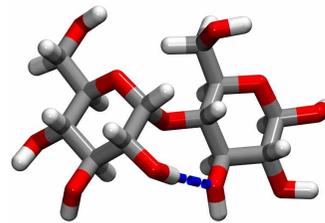
Disaccharides



Trehalose



Sucrose



Maltose

⊕ Same chemical formula ($C_{12}H_{22}O_{11}$)

⊕ Same number of OH groups (8)

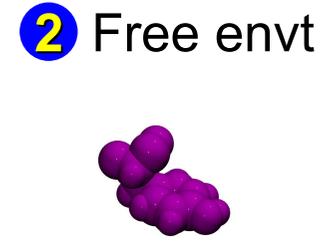
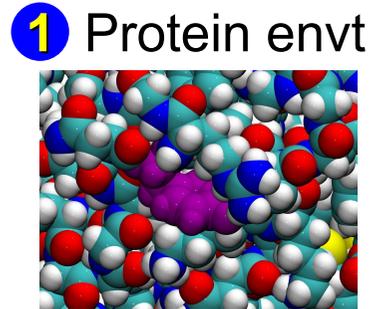
⊕ Different topologies

MD program	Force fields	Concentrations	T (K)	Δt	t_{sim}
CHARMM	<p>W SPC/E</p> <p>D CSFF</p> <p>L CHARMM22</p>	0, 37, 50, 60 wt % protein-free basis (~ 14-16 katoms)	300	2 fs	10-17 ns

Fractional solvent accessibilities

$$f_{sa} = \frac{SASA_{res,protein}}{SASA_{res,free}}$$

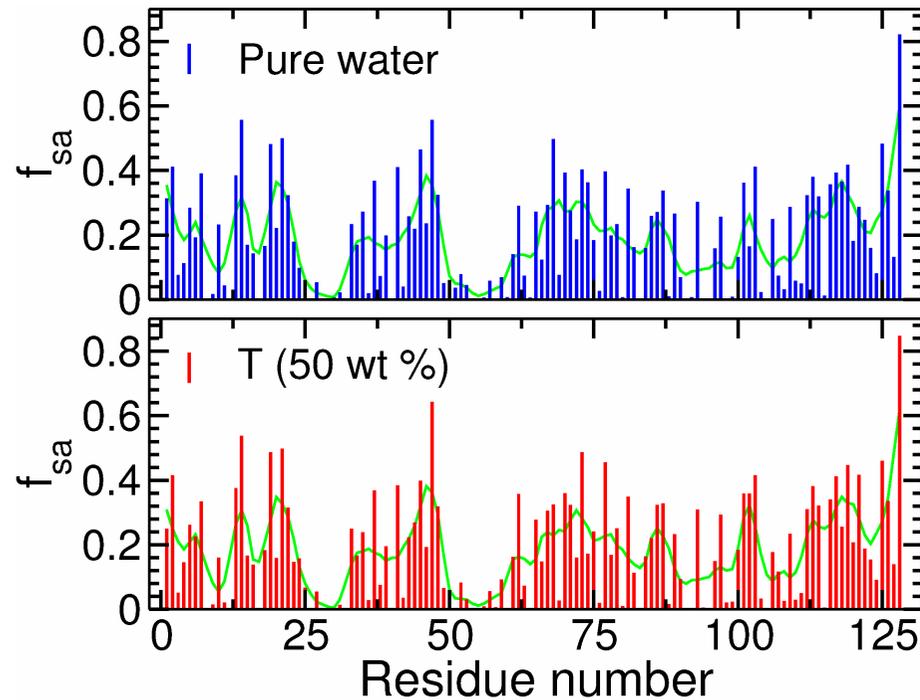
Ex.: Tyr20



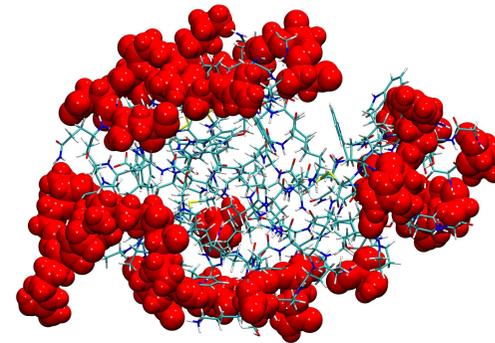
1 SASA = 70 Å²

2 SASA = 347 Å²

➡ $f_{sa} = 0.20$

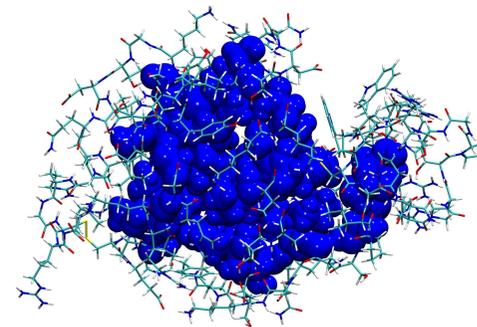


30 most exposed residues :

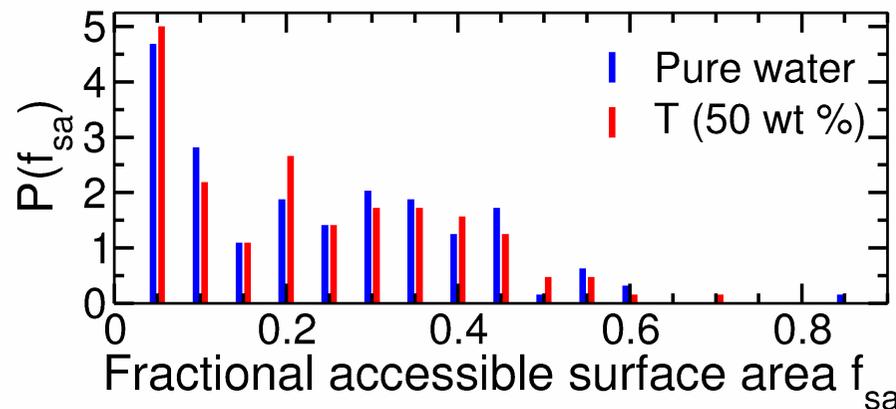


Surface residues

30 least exposed residues :

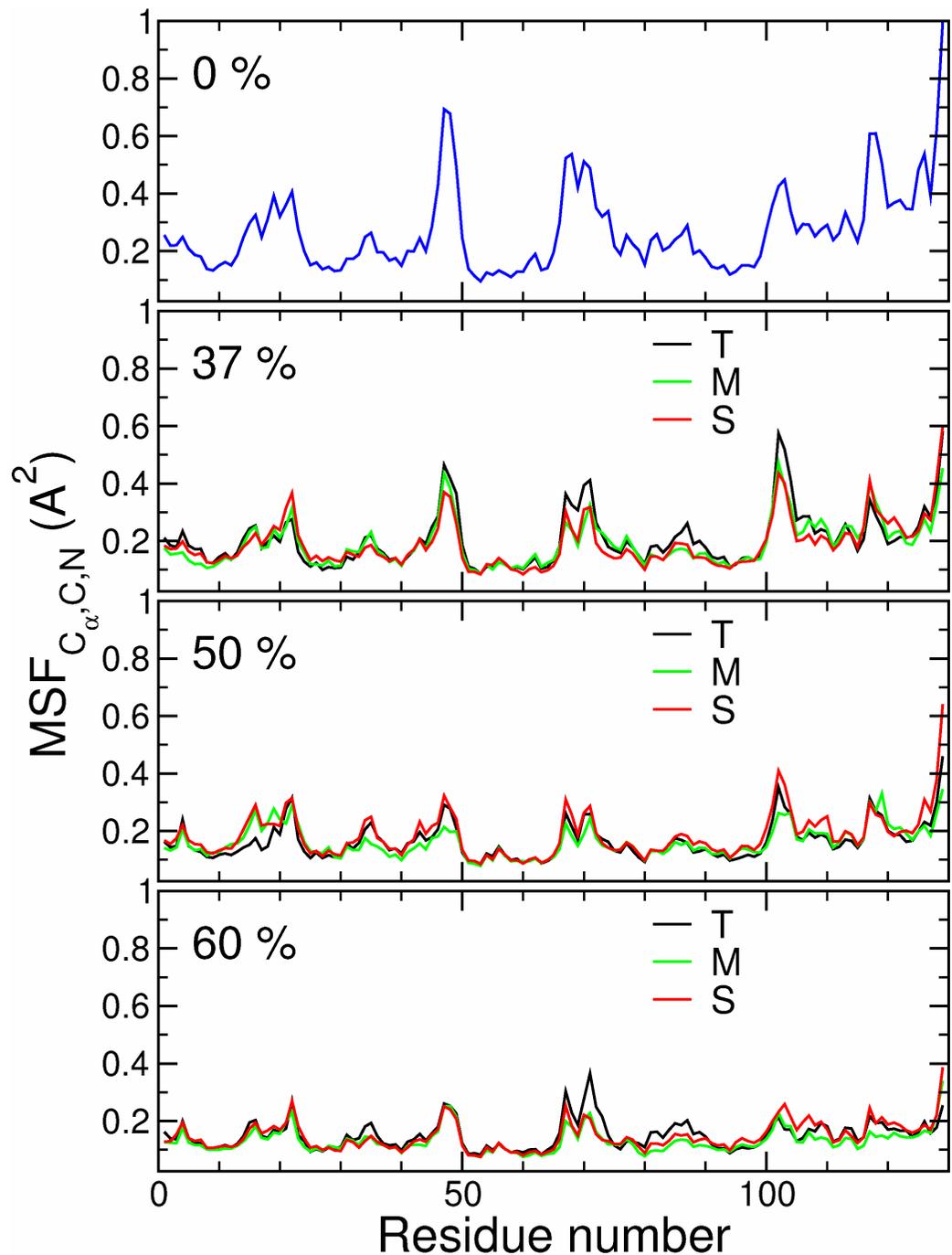


Core residues



➡ No effect of sugars on the native conformation of lysozyme

Mean-square fluctuations $\langle u^2 \rangle$



■ High heterogeneity of local environments

➡ similar to residues f_{sa}

■ Addition of sugars

⊕ significant reduction of $\langle u^2 \rangle$

➡ high T° -shift of the denaturation temperature T_m

⊕ No change of the global shape

➡ no preferential interaction with specific parts

⊕ Similar effect on lysozyme

Local concentration of water around lysozyme

$$g_{N,Ow}(r) = \frac{\frac{n_{Ow}(r)}{n_{Ow}(r)+n_{Os}(r)}}{\frac{N_{Ow}}{N_{Ow}+N_{Os}}}$$

n/N : local/total numbers of atoms

r : minimal distance to any heavy atom of lysozyme

■ $g_{N,Ow} > 1$ ➡ excess of water

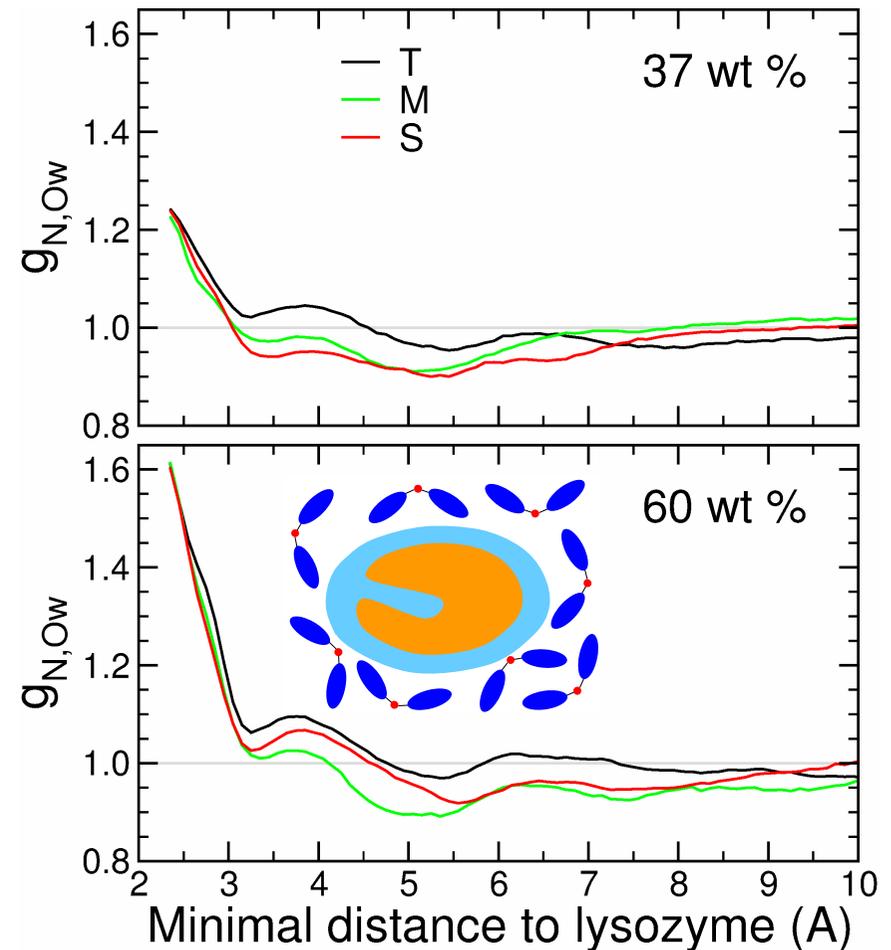
■ $g_{N,Ow} < 1$ ➡ lack of water

➡ preferential exclusion of sugars

➡ agreement with the *preferential hydration* hypothesis [Timasheff, *Biochem.* 41 (2002),89]

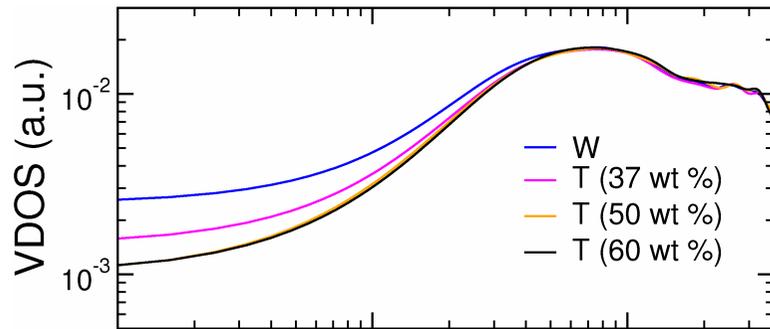
■ T : slightly more excluded ➡ larger hydration number n_H (T: 7.95, M : 6.50, S: 6.33)

[Kawai et al., *Cryobiol.* 29 (1992), 599]



Vibrational density of states (1)

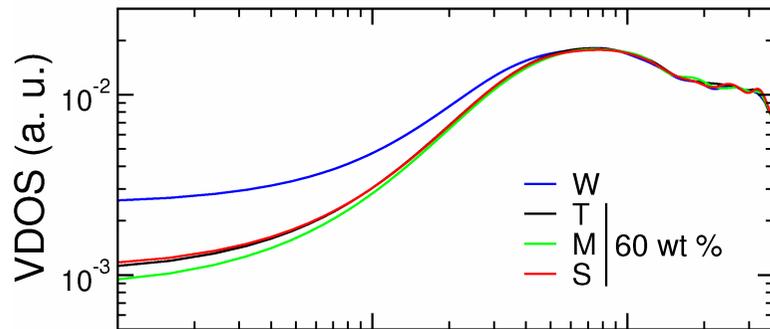
Lysozyme



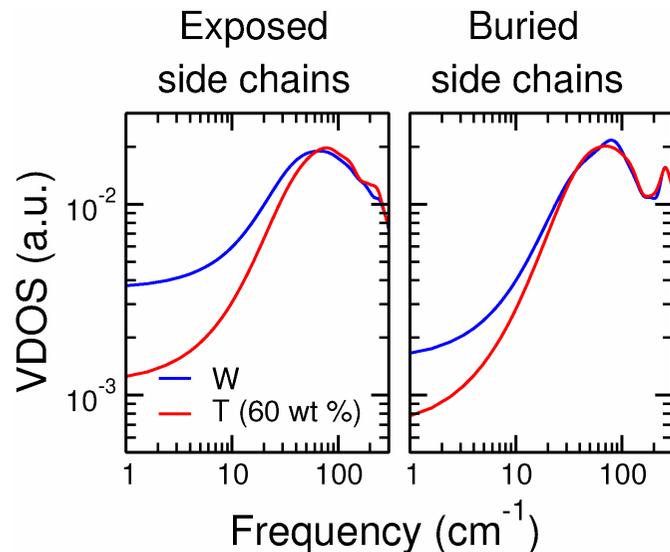
■ Decrease of the VDOS with sugar concentration

➡ more rigid lysozyme

➡ stabilization at high temperatures



➡ Small differences between sugars, within simulation accuracy



■ Reduced VDOS for buried residues

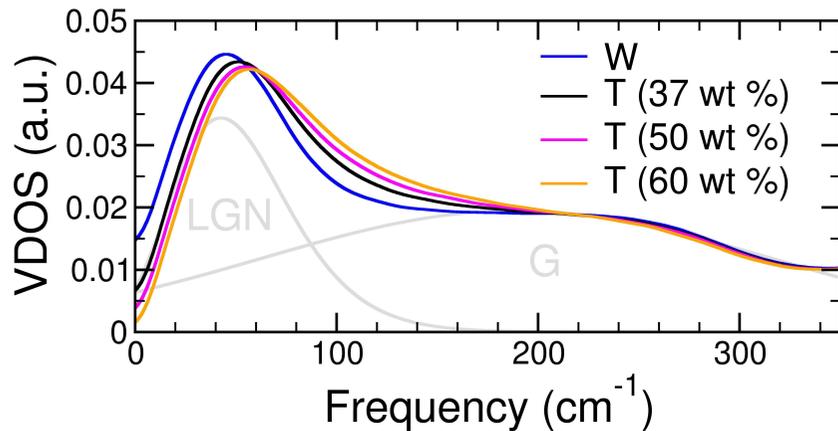
➡ lower mobility, in line with $\langle u^2 \rangle$

■ Lower effect of sugars on buried residues

➡ indirect influence

Vibrational density of states (2)

Water

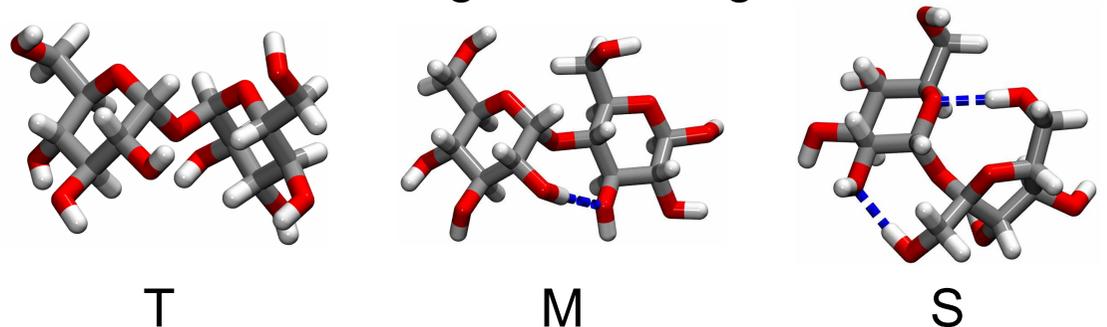


- Broadening and high freq. shift of the 1st peak
- ➡ Increased **heterogeneity** and **rigidity** of water local environments

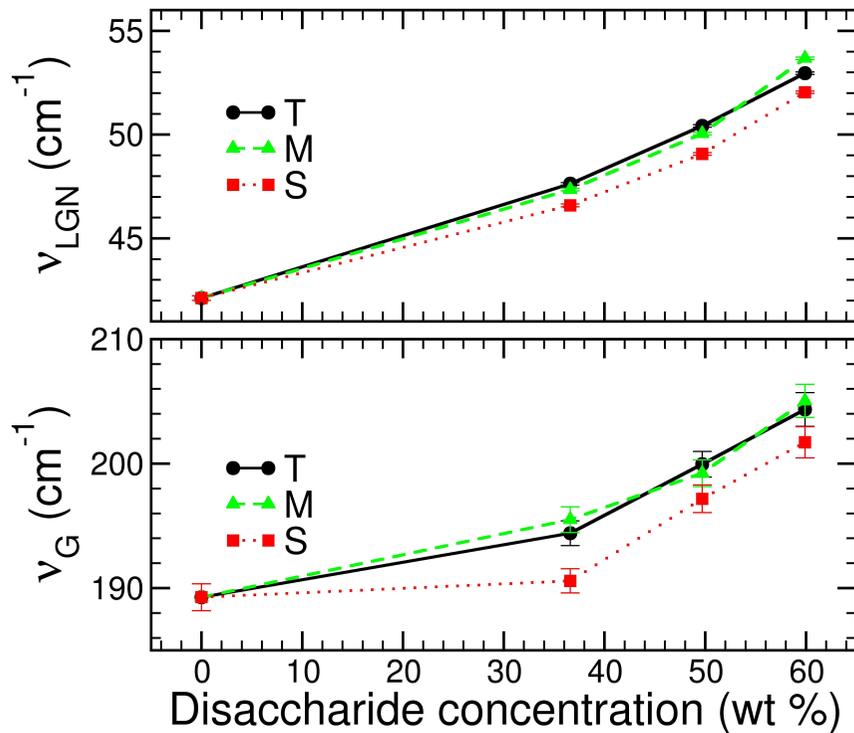
- Fit with a log-normal (LGN) and a gaussian (G) functions :

- ➡ strengthening of the water HB network

- ➡ $T \sim M > S$
 - ⬢ Lower nb of sugar-water HBs
 - ⬢ Larger nb of sugar intra HBs



- Differences between sugars too small to be observed on the protein VDOS



Relaxation properties

$\tau = 1/e$ decay time of the incoherent intermediate scattering function $S_{inc}(Q=2.29 \text{ \AA}^{-1}, t)$

☑ Distributions similar to f_{sa} and $\langle u^2 \rangle$

➡ $\tau_{core} > \tau_{surface}$ (as expected)

☑ Significant slowing down of lysozyme's :

⊕ Global translation/rotation

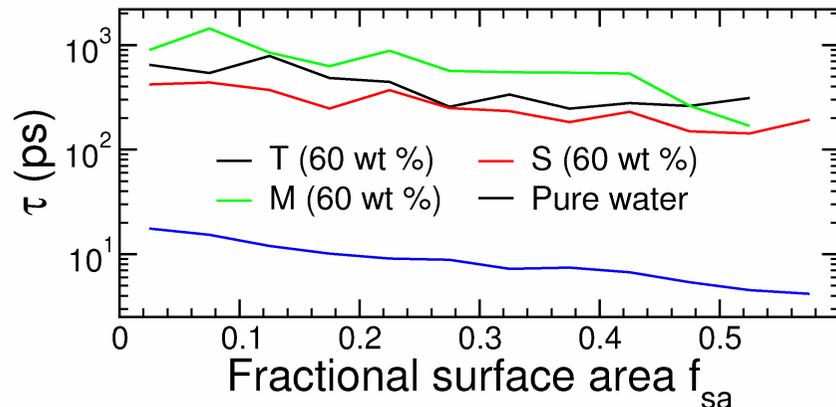
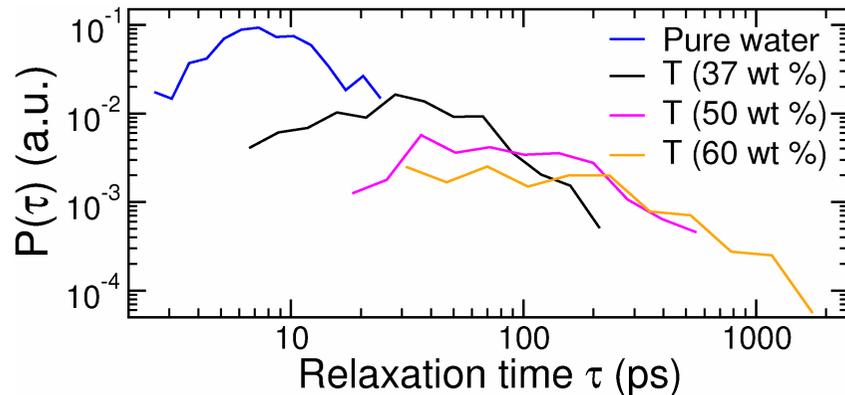
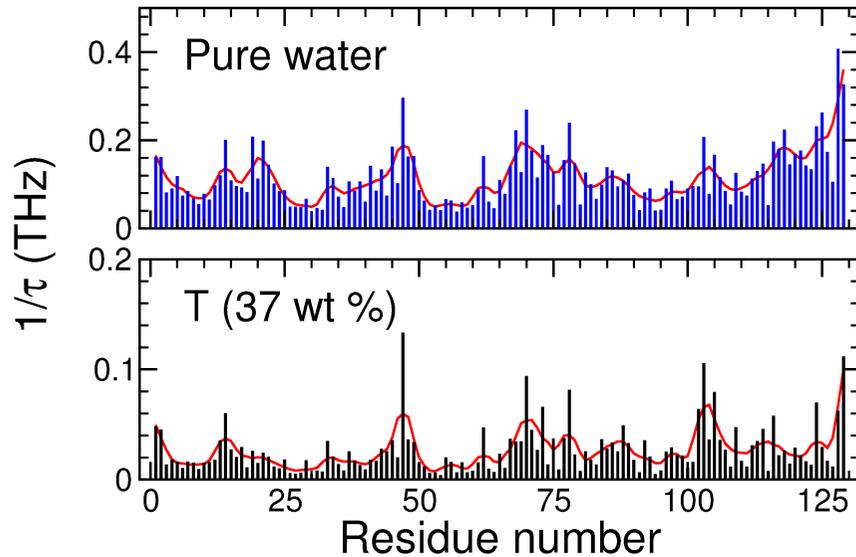
⊕ Internal dynamics ($\langle u^2 \rangle$)

☑ On average :

■ $\log(\tau) \sim f_{sa}$

■ $\tau(M) > \tau(T) > \tau(S)$

➡ sugar cluster size $\langle n_s \rangle$: $M > T > S$



Conclusions

➤ Influence of sugars on lysozyme

- ☑ none on its native conformation → stabilizing solutes
- ☑ significant reduction of $\langle u^2 \rangle$ → shift of its heat denaturation temperature T_m
- ☑ preferential exclusion of sugars → non-specific interaction
- ☑ strengthening of the water HB network → protein-solvent dynamical coupling via its water hydration shell
- ☑ more important for exposed than for buried residues

➤ Comparison between sugars

- ☑ rather similar effects on native lysozyme



Similar effects on denatured lysozyme ? →

MD simulations of denatured lysozyme needed

Selected papers

- *Analysis of sugar bioprotective mechanisms on the thermal denaturation of lysozyme from Raman scattering and differential scanning calorimetry investigations*, A. Hédoux, J.-F. Willart, R. Ionov, F. Affouard, Y. Guinet, L. Paccou, A. Lerbret, M. Descamps, *J. Phys. Chem. B* **2006**, 110, 22886-22893.
- *How do trehalose, maltose, and sucrose influence some structural and dynamical properties of lysozyme ? Insight from molecular dynamics simulations*, A. Lerbret, P. Bordat, F. Affouard, A. Hédoux, Y. Guinet, M. Descamps, *J. Phys. Chem. B* **2007**, 111, 9410-9420.
- *Molecular dynamics simulations of lysozyme water/sugar solutions*, A. Lerbret, F. Affouard, P. Bordat, A. Hédoux, Y. Guinet, M. Descamps, *Chem. Phys.* **2008**, 345, 267-274.
- *Low-frequency vibrational properties of lysozyme in sugar aqueous solutions: a Raman scattering and molecular dynamics simulation study*, A. Lerbret, F. Affouard, P. Bordat, A. Hédoux, Y. Guinet, M. Descamps, *J. Chem. Phys.* **2009**, 131, 245103.